

Supplementary Information

High Resolution Data Independent Acquisition Electron Transfer Dissociation Mass Spectrometry: Multiplexed Analysis of Post-translationally Modified Proteins

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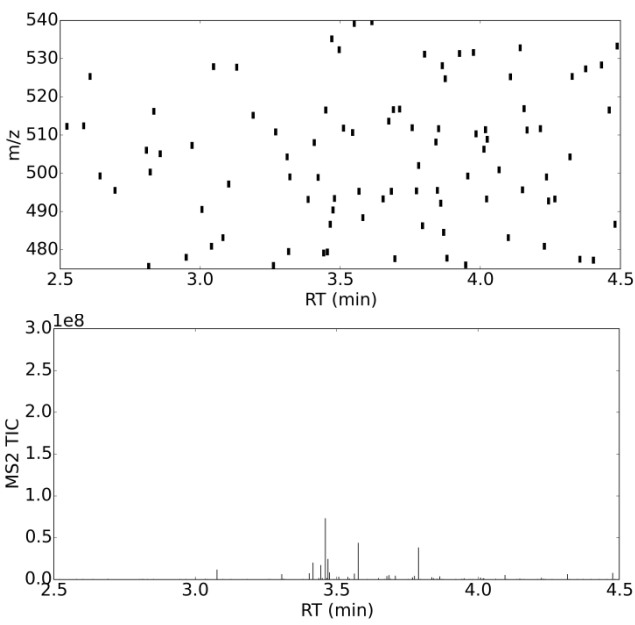
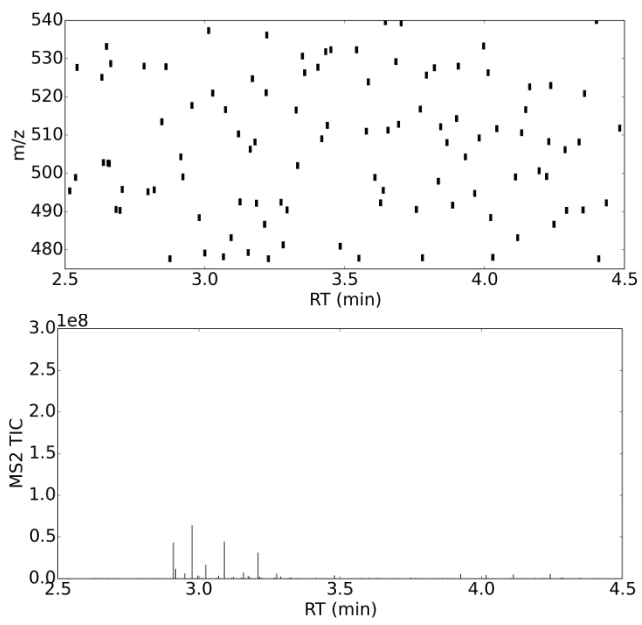
S1

A

DDA

Rep 1

Rep 2



B

DIA

Rep 1

Rep 2

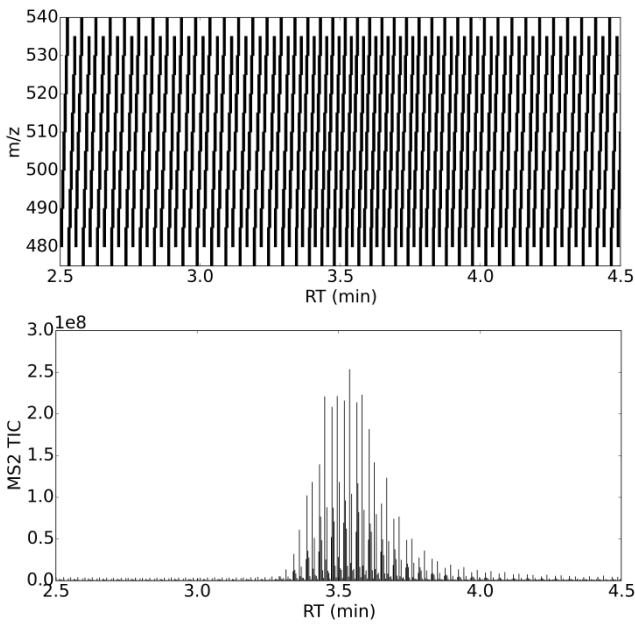
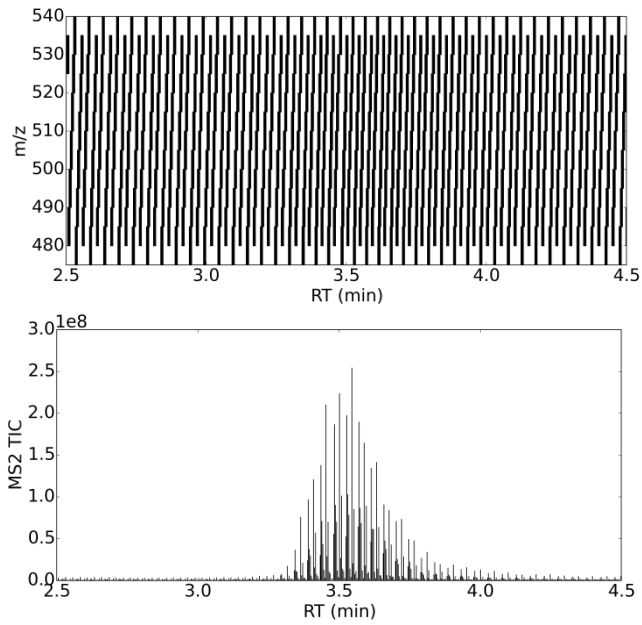


Figure S1: Shown are two replicate analyses in (A) DDA ETD and (B) DIA ETD mode. (A) DDA shows sporadic sampling over the selected mass range (top) resulting in unreliable MS2 total ion chromatograms that vary significantly from run to run (bottom, compare left and right). The bars in the top panel represent actual isolation windows at given retention times. (B) DIA shows predictable and reliable sampling over the selected mass range (top) resulting in reproducible MS2 total ion chromatograms that are identical from run to run (bottom, compare left and right). The bars in the top panel represent actual isolation windows at given retention times.

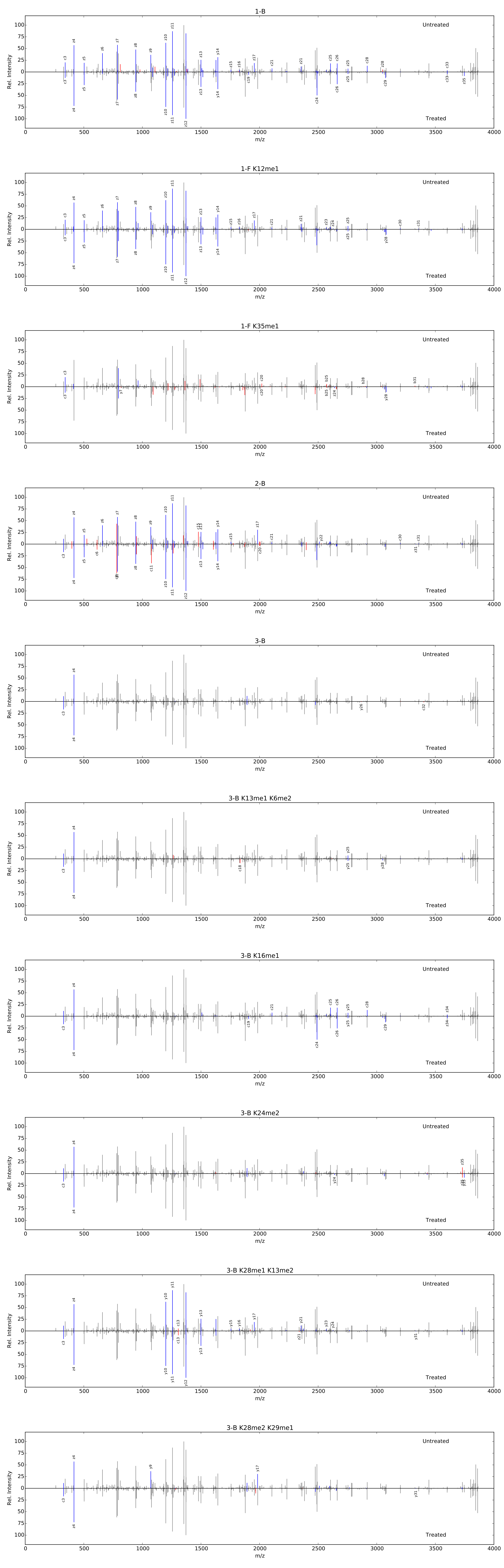


Figure S2: Displayed are the fragment ion matches of the minimum set of proteoforms (1-B, 1F, K12me1, 1F K35me1, 2-B, 3-B, 3-B K13me1 K6me2, 3-B K16me1, 3-B K24me2, 3-B K28me1 K13me2, and 3-B K28me2 K29me1) for the 475-485 precursor ion windows of the untreated and treated samples. Ions highlighted in blue are shared by multiple proteoforms, ions highlighted in red are unique to a particular proteoform. H2B3BK16me1 was a proteoform that was deduced to be present because of its precursor ions indicating a methylated H2B3B.

| # Explained Fragment Ions | Replicate 1 | Replicate 2 |
|---------------------------|-------------|-------------|
| CID | 37 | 30 |
| ETD | 115 | 117 |

Table S1: Comparison of CID and ETD in DIA mode shows ETD is superior to CID in the number of explained fragment ions. ETD produced approximately three times more fragment ions that could be assigned to the H2B proteoforms present. This difference was significant with a p-value of 0.002.